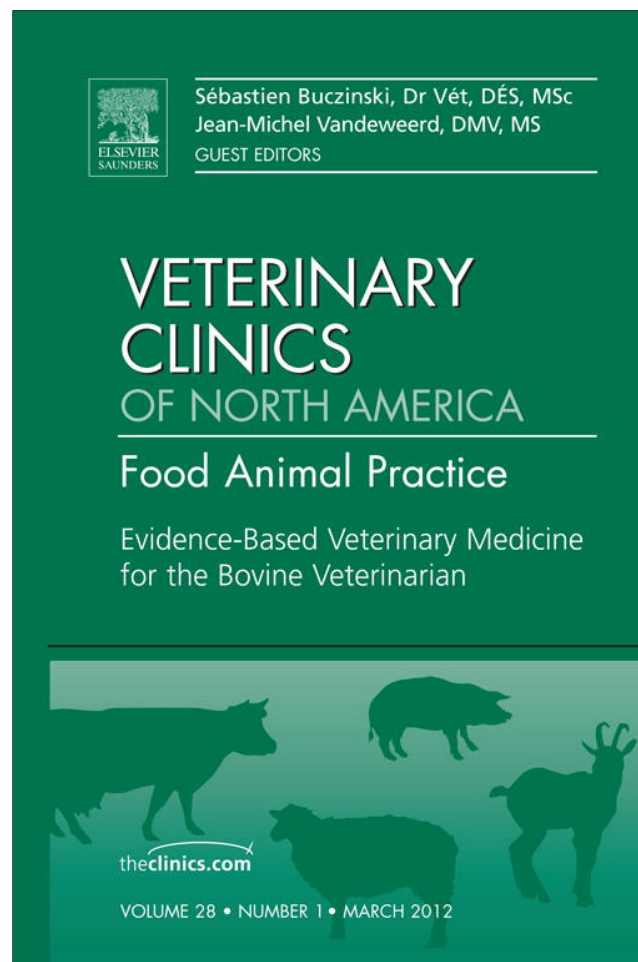


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Evidence-Based Effectiveness of Vaccination Against *Mannheimia haemolytica*, *Pasteurella multocida*, and *Histophilus somni* in Feedlot Cattle for Mitigating the Incidence and Effect of Bovine Respiratory Disease Complex

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KEYWORDS

- Evidence-based • Bovine respiratory disease complex
- *Mannheimia haemolytica* • *Pasteurella multocida*
- *Histophilus somni* • Vaccination

Evidence-based medicine (EBM) was introduced to the medical literature in a 1992 article by the Evidence-based Working Group at McMaster University Health Sciences Centre in Canada to describe the clinical learning strategy they had been developing for over a decade.¹ The principles of EBM are being applied to the veterinary profession under the term “evidence-based veterinary medicine” (EBVM).^{2–4} The underlying concepts of EBM and EBVM are rooted in clinical epidemiology and are not new but represent a

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formal and explicit effort to increase the occurrence of basing clinical decisions on a dispassionate review of published trials that adequately meet a *priori* standards of experimental design and experimental execution.

Although most clinical decisions in veterinary medicine are based on evidence of some type, some evidence is very strong (rigorously tested in the target species under natural conditions, such as cattle in commercial feedlots in experiments designed to prove a theory to be false), some evidence is very weak (not tested), and some is intermediate.⁵⁻⁷ The hierarchy of evidence is based on the strength of evidence for causation, the ability of the study to control bias, and the similarity between the study population and the population currently being considered in a clinical setting.

With respect to bacterial vaccination in feedlot cattle, sources regarded as the strongest evidence for the effectiveness of vaccination against *Mannheimia haemolytica*, *Pasteurella multocida*, and *Histophilus somni* for mitigating the incidence and effect of bovine respiratory disease (BRD) complex are randomized controlled clinical trials in feedlot cattle under a typical husbandry environment with adequate blinding of investigators, a clear case-definition of BRD, and adequate intensity and length of follow-up; or systematic reviews of more than 1 trial that meet these criteria. In addition, other available evidence, including studies testing the effects of vaccination of cattle exposed to pathogen-challenged disease models, studies testing the effects of vaccination of cattle in dissimilar production settings (ie, dairy calves), and studies using in vitro methodologies to test vaccination effects can be used as indirect indicators in the clinical decision-making process, particularly when higher levels of evidence are lacking.

The “body of evidence” for this clinical question is the sum of multiple studies investigating the effect of vaccines against *M haemolytica*, *P multocida*, and *H somni* administered to cattle. Each individual research study contributes to that body of evidence and each publication can be ranked on a scale from weak evidence to very strong evidence, which, for the veterinary practitioner, implies an increasing confidence in recommendations based on a particular study. And, although a simple ranking of experimental trial types is helpful to describe ascending levels of evidence, by its simplistic nature, it incorrectly depicts levels of evidence as a one-dimensional and straightforward hierarchy. For example, veterinarians are often confronted with determinations such as—Which is better evidence, a randomized trial in 3 month-old dairy calves (ie, nontarget animals, but a study design with high control of bias and confounding) or a pathogen-challenged disease model study in feedlot cattle (ie, study with less external validity but in the target population)? In these situations, the clinical expertise, experience, and judgment of the veterinarian must be used to aid the ranking of evidence generated by these studies and to guide recommendations for use of bacterial respiratory pathogen vaccines into processing protocols in the field.

Veterinarians considering the strength of evidence must use several perspectives to determine the reliability of research for clinical use.

1. The first consideration is the internal validity of the research, which is determined by the study method and appropriate use of controls for bias. Research reports with good internal validity provide assurance that the results represent an unbiased estimate of the true direction and magnitude of the treatment effect in the study population. For randomized controlled studies, accepted methods of random allocation and blinding of study investigators to the treatment for each experimental unit are key experimental design features to avoid bias and confounding.

2. The second consideration is the population used in the research and its appropriateness as a model for the population that generated the clinical question. Generally, the target species in similar housing and husbandry environments provides stronger evidence than the target species in significantly different housing and husbandry environments, related species, unrelated species, or in vitro methods.
3. And, third, the clinical relevance of the outcomes of the research should be considered with patient- or herd-oriented outcomes (eg, morbidity risk, mortality risk, or average daily weight gain) providing more direct evidence of intervention effectiveness than disease-oriented outcome measurements such as behavior frequency, body temperature, or antibody response.

Using these considerations, the highest rating in all 3 dimensions would provide the highest level of evidence.

MATERIALS AND METHODS

A literature search was conducted to identify studies published in English that reported the effectiveness of *Mannheimia (Pasteurella) haemolytica*, *Pasteurella multocida*, and *Histophilus (Haemophilus) somnus* vaccination in cattle. A search strategy using (*Mannheimia haemolytica* OR *Pasteurella haemolytica* OR *Pasteurella multocida* OR *Haemophilus somnus* OR *Histophilus somni*) AND (respiratory disease OR pneumonia OR pneumonic) AND (bovine OR cattle OR bos) AND (vaccine OR vaccinate) was used to query PubMed (n = 164 references), CAB abstracts (n = 379) and Biologic Abstracts (n = 160) followed by a hand search through cited references (n = 4). A published manuscript is considered a “study” while a “trial” is a direct comparison of a vaccine treatment to a control treatment within a study. A single study may include more than 1 trial. After reading the abstract from each unique publication, 34 studies were included in this review. Fifteen studies (22 trials) were considered the highest level of evidence in that they were trials using feedlot or stocker cattle in North American production settings appropriately allocated to treatment groups with naturally occurring disease.^{8–22} One or more trials from 5 other studies were identified that used feedlot cattle in typical North American production settings, but they were weakened by lack of blinding, treatment being confounded by arrival group or other vaccine treatment, or significant loss to follow-up and were discarded from the summary.^{23–27} In addition, 3 terminal studies (5 trials) investigated the use of commercially available vaccines in feedlot cattle with a pathogen-challenged disease model^{14,28,29}; 3 studies (5 trials) used dairy or beef calves with naturally occurring disease to investigate effects of vaccination^{27,30,31}; and 13 studies investigated the use of commercially available vaccines in dairy calves with an induced-disease model.^{32–44} Studies were excluded from the review if they did not report original data (primary study); if they did not include a nonvaccinated/placebo control group; if the outcome did not include an assessment of morbidity risk, mortality risk, or extent of lung involvement (eg, only reported serologic titers); or if the same results were published in a more complete form elsewhere. Many studies did not report specific allocation schemes used or whether effective blinding occurred, and some studies used inappropriate statistical tests for the data collected. Studies with obvious limitations due to experimental design were excluded, but studies with poorly described experimental designs were retained.

A meta-analysis was performed, and a Mantel-Haenszel risk ratio (RR) and 95% confidence interval (95% CI) were calculated for each trial reporting cumulative incidence of BRD morbidity or mortality (or crude morbidity or mortality).⁴⁵ Calculated

RR less than 1.0 indicates that vaccinates had lower cumulative incidence compared to controls, while RR greater than 1.0 indicates that vaccinates had higher cumulative incidence compared to controls. In order to be considered to have a statistically significantly lower morbidity or mortality cumulative incidence in vaccinates compared to controls, the upper limit of the 95% CI must be below 1.0; while in order to consider the cumulative incidence of morbidity or mortality to be statistically significantly higher in vaccinates compared to controls, the lower limit of the 95% CI must be greater than 1.0. A Forest plot is provided to demonstrate graphically the relative strength of the treatment effects.

RESULTS

Studies Using Feedlot Cattle With Naturally Occurring Disease (Appendix 1)

Data were extracted from the 15 studies (22 trials) that tested the effectiveness of vaccination against 1 or more of the bacterial pathogens *M haemolytica*, *P multocida*, and *H somni* in feedlot cattle for mitigating the incidence and effect of BRD complex using feedlot cattle with naturally occurring disease in order to calculate the RR for each trial (Appendix 1). Using the criteria outlined here, these studies are expected to provide the highest level of evidence from the available studies identified in the literature search. A brief account of the studies, including a description of how the cattle were allocated to treatment, the timing of vaccine administration, and a characterization of the vaccines used, can be found in the appendices.

All 22 trials reported a cumulative incidence for morbidity. For some trials the case definition for being considered a case was not specified; other studies had clear case definitions for BRD morbidity. Some studies reported crude morbidity and mortality risk (morbidity or mortality due to any cause), while some studies reported BRD-specific morbidity and mortality risk.

M haemolytica and M haemolytica + P multocida vaccines

Studies investigating the effectiveness of several different commercially available vaccines against *M haemolytica* (15 trials) and *M haemolytica* + *P multocida* (3 trials) were summarized, with 3 of 18 trials reporting a statistically significant reduction in BRD morbidity cumulative incidence in vaccinates compared to controls (eg, upper 95% CI was less than 1.00),^{10,16,17} while 4 reported an increased risk of BRD morbidity^{8,17,20} and 11^{9–15,18–20} reported a decreased risk of BRD morbidity cumulative incidence that was not different from control populations (Fig. 1). The summary RR for these trials is 0.93 with a 95% CI that does not cross 1.0 (0.89–0.98), indicating a statistically significant lower risk of morbidity in vaccinated feedlot cattle compared to controls.

The 15 trials that investigated the effect of *M haemolytica*-only vaccine accounted for 90% of the weighted summary RR, and 2 of 15 trials reported a statistically significant reduction in BRD morbidity cumulative incidence in vaccinates compared to controls,^{16,17} while 3 reported an increased risk of BRD morbidity^{17,20} and 10^{9–15,18–20} reported a decreased risk of BRD morbidity cumulative incidence that was not different from controls. The 3 trials that investigated the effect of *M haemolytica* + *P multocida* vaccination accounted for 10% of the weighted summary RR. One of the 3 trials reported a statistically significant reduction in BRD morbidity cumulative incidence in vaccinates compared to controls,²³ while 1 reported an increased risk of BRD morbidity⁸ and 1 reported a decreased risk of BRD morbidity cumulative incidence that was not different from control populations.²³

Evaluating mortality RR in 9 studies that measured BRD-specific or crude mortality risk indicates that 7 trials reported decreased cumulative mortality incidence that was

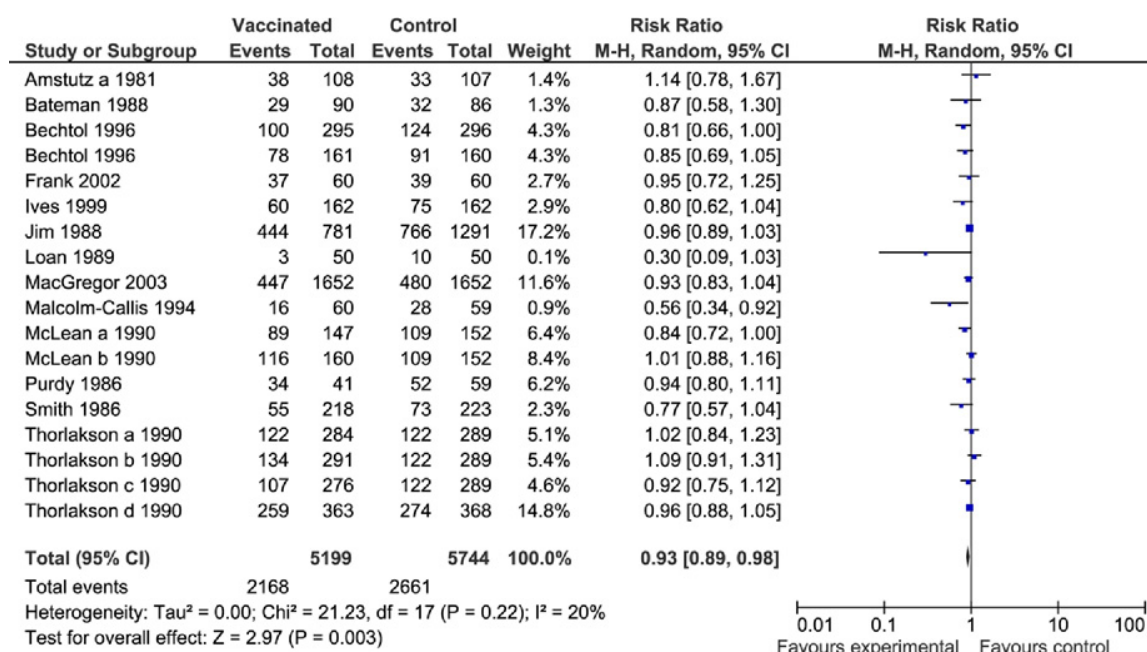


Fig. 1. Forest plot of RR for 18 trials comparing cumulative morbidity incidence of feedlot cattle vaccinated against *M haemolytica* (15 trials) or *M haemolytica* + *P multocida* (3 trials) compared to controls.

not different in vaccinates relative to controls, while 2 reported an increased risk of mortality that was not different from control populations.^{10,12,15–18,20} An additional 6 trials reported cumulative mortality incidence, but the RR could not be calculated because of nonevents (zero for very low count cells) (**Fig. 2**). The summary RR for these trials is 0.76 with a 95% CI that crosses 1.0 (0.56–1.04), indicating mortality risk in vaccinated feedlot cattle is not statistically different than that of controls.

M haemolytica + *H somni* vaccine studies

One study investigated the effectiveness of a commercially available vaccine against *M haemolytica* + *H somni* in feedlot cattle with natural disease challenge.²¹ In this study, vaccinated cattle had statistically significantly lower morbidity compared to controls. There were no deaths in the vaccinates or controls (**Appendix 1**).

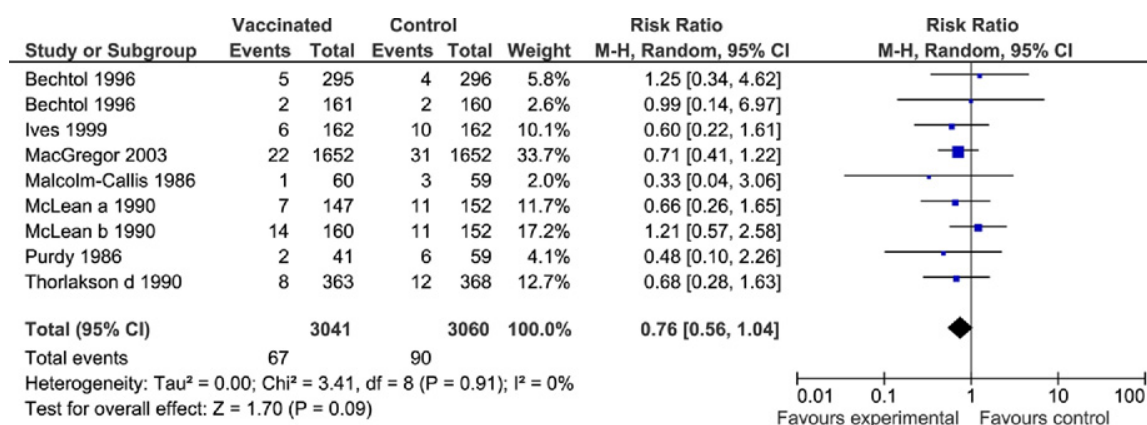


Fig. 2. Forest plot of RR for 9 trials comparing cumulative mortality incidence of feedlot cattle vaccinated against *M haemolytica* (7 trials) or *M haemolytica* + *P multocida* (2 trials) compared to controls.

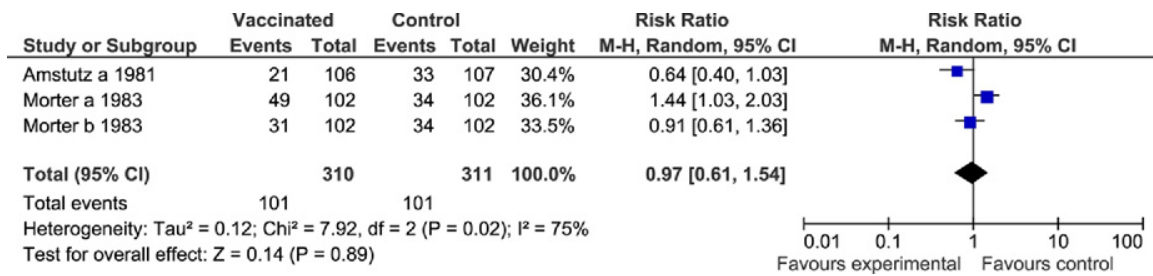


Fig. 3. Forest plot of RR for 3 trials comparing cumulative morbidity incidence of feedlot cattle vaccinated against *H somni* to controls.

H somni vaccine studies

Three trials were identified that investigated the effectiveness of *H somni* vaccination of feedlot cattle to decrease the cumulative incidence of BRD due to natural challenge.^{8,22} The summary RR is 0.97 (95% CI, 0.61–1.54), indicating that BRD morbidity risk of vaccinated cattle was not statistically different than controls (**Fig. 3**).

As these studies provide the highest level of evidence for making the clinical decision about the effectiveness of vaccination against the pathogens *M haemolytica*, *P multocida*, and *H somni* in feedlot cattle for mitigating the incidence and effect of BRD complex, the weight of evidence from these 22 trials is particularly important. The summary RR indicates that these studies indicate that vaccination against *M haemolytica* or *M haemolytica* + *P multocida* has the potential to decrease the incidence of BRD complex in feedlot cattle, but the numerical decrease in mortality risk was not statistically different from controls. Much less evidence is available to determine the effectiveness of vaccination against *H somni* in feedlot cattle, and although these studies using natural disease challenge indicate that the risk of BRD does not appear to be affected by vaccination against this pathogen, we have very little power to detect a true difference if it did exist.

Studies Using Feedlot Cattle With Pathogen-Challenged Disease Models (Appendix 2)

M haemolytica vaccines

Three studies reporting 5 trials were identified that used feedlot cattle to evaluate the association between vaccination with commercially available *M haemolytica* vaccines and mortality risk and lung lesion severity following induced disease with a transthoracic inoculation of *M haemolytica*.^{14,28,29} All 5 trials reported increased survival post challenge, and the 4 trials that reported lung severity indicated decreased percentage of total lung volume being classified as pneumonic in vaccinates compared to controls.

Studies Using Dairy or Beef Calves With Naturally Occurring Disease (Appendix 3)

M haemolytica and *M haemolytica* + *P multocida* vaccines

Studies using dairy or beef calves during the first 3 to 6 months of life to test the efficacy of a vaccine against *M haemolytica* or a combination vaccine against *M haemolytica* + *P multocida* are not considered to provide a high level of evidence for clinical questions arising from feedlot cattle health problems because of differences in age, housing, and management. **Figure 4** depicts the Forest plots of the RR for BRD morbidity for 3 trials using dairy calves vaccinated against *M haemolytica* (2 trials) or *M haemolytica* + *P multocida* (1 trial).^{27,30} **Figure 5** depicts the Forest plot of the RR for crude mortality for 2 dairy calf trials evaluating *M haemolytica* vaccine.²⁷

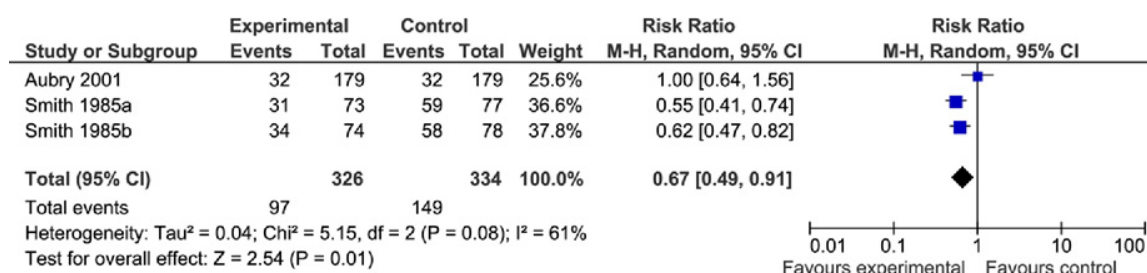


Fig. 4. Forest plot of RR for 3 trials comparing cumulative morbidity incidence of dairy calves vaccinated against *M haemolytica* or *M haemolytica* + *P multocida* compared to controls.

The trials that evaluated the effectiveness of *M haemolytica* or *M haemolytica* + *P multocida* revealed summary RR indicating a statistically significant reduction in BRD morbidity (**Fig. 4**), but not crude mortality (**Fig. 5**) in vaccinated calves compared to controls.

M haemolytica + *H somni* vaccine studies

Calves vaccinated with a genetically attenuated leukotoxin of *M haemolytica* combined with bacterial extracts of *M haemolytica* and *H somni* did not have statistically significantly different risk of BRD morbidity compared to controls (**Fig. 6**).³¹

SUMMARY

The clinical question of whether to use commercially available vaccines against bacterial pathogens associated with BRD in feedlot cattle is important to the veterinarians and producers making the decision, as well as to the health and well-being of feedlot cattle. Making an evidence-based clinical decision based primarily on published, scientifically accepted controlled trials using feedlot cattle, with supportive information from published trials using pathogen-challenged disease models or using dairy or beef calves housed and managed under different husbandry systems, requires not only the gathering and summarizing of the available information but also considering the context of specific clinical questions.

The summary data would indicate potential benefit for vaccination of feedlot cattle against *M haemolytica* and *P multocida* with no evidence of benefit for vaccination against *H somni* for mitigating the incidence and effect of BRD complex. Unfortunately, the published body of evidence does not provide a consistent estimate of the direction and magnitude of effectiveness in feedlot cattle vaccination against *M haemolytica*, *P multocida*, or *H somni*.

One limitation for the conclusions that can be drawn from this group of studies includes the fact that all the feedlot studies with natural disease challenge mixed

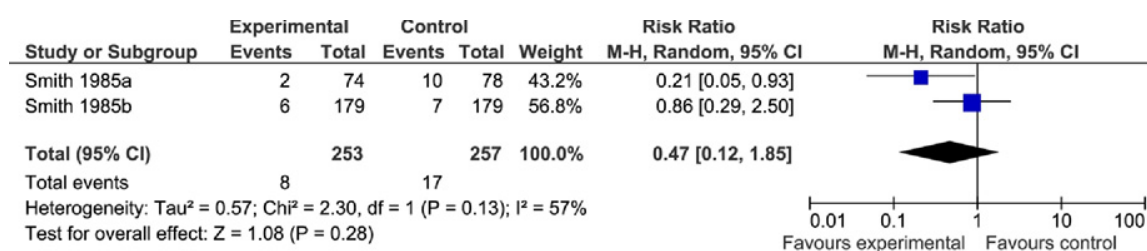


Fig. 5. Forest plot of RR for 2 trials comparing cumulative mortality incidence of dairy calves vaccinated against *M haemolytica* compared to controls.

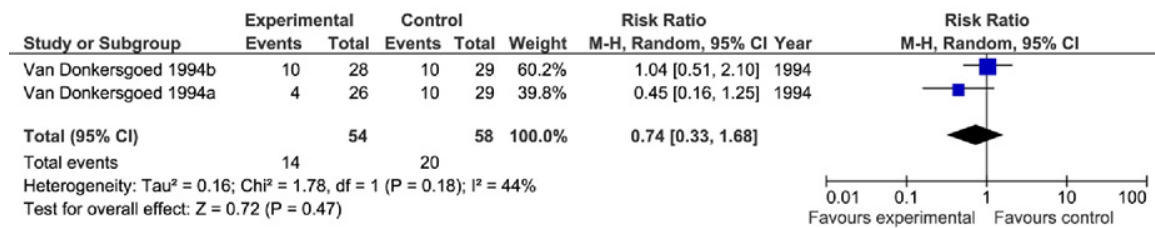


Fig. 6. Forest plot of RR for 2 trials comparing cumulative morbidity incidence of dairy calves vaccinated against *M haemolytica* + *H somni* compared to controls.

vaccinated and unvaccinated calves in the same feedlot pens. This mixing may underestimate the value of vaccination because of the phenomena of herd immunity. In mixed pens, the vaccinated calves may reduce the disease challenge for unvaccinated controls and unvaccinated calves may increase the disease challenge for vaccinated calves compared to the exposure expected when entire pens are either vaccinated or not vaccinated. Another limitation is that some studies reported crude morbidity and mortality while other studies reported BRD-specific morbidity and mortality. Approximately 59% of the weighted summary RR for morbidity in the feedlot studies was derived from studies using a case definition for BRD as the criteria for being classified as a morbid animal, while 41% of the weighted summary RR came from studies reporting the effect of vaccination in all causes of morbidity. Similarly, approximately 57% of the weighted summary RR for mortality in the feedlot studies came from studies specifying mortalities associated with BRD, while 43% of the weighted summary RR was derived from studies reporting the effect of vaccination on all causes of mortality. If non-BRD mortalities were evenly distributed between vaccinates and controls in these studies, aggregating mortality of all causes to test the association with vaccination status will decrease the RR between vaccinates and nonvaccinated controls.

A thorough search of the published literature and a structured meta-analysis to produce a summary Mantel-Haenszel RR and 95% CI are helpful tools for making an assessment of the evidence for the effectiveness of vaccination against *M haemolytica*, *P multocida*, and *H somni* for mitigating the incidence and effect of BRD complex in feedlot cattle. However, because of the limitations of the studies used in the meta-analysis and the various specific clinical situations that feedlot veterinarians and producers confront, it is necessary to combine this summary with other sources of information and unpublished data, as well as continued monitoring of recommendations to arrive at the best advice for feedlot clients.

REFERENCES

1. Evidence-Based Medicine Working Group. Evidence-based medicine: a new approach to teaching the practice of medicine. *JAMA* 1992;268:2420–5.
2. Keene BW. Towards evidence-based veterinary medicine [editorial]. *J Vet Intern Med* 2000;14:118–9.
3. Doig GS. Evidence-based veterinary medicine: what it is, what it isn't and how to do it. *Aust Vet J* 2003;81:412–5.
4. Roudebush P, Allen TA, Dodd CE, et al. Evidence-based medicine: applications to veterinary clinical nutrition. *J Am Vet Med Assoc* 2004;224:1766–71.
5. Sackett DL. Rules of evidence and clinical recommendations. *Can J Cardiol* 1993;9:487–9.
6. Dans AL, Dans LF, Guyatt GH, et al. Users' guide to the medical literature. XIV. How to decide on the applicability of clinical trial results to your patient. *JAMA* 1998;279:545–9.

7. Berg AO. Dimensions of evidence. In: Geyman JP, Deyo RA, Ramsey SD, editors: Evidence-based clinical practice: concepts and approaches. Boston: Butterworth-Heinemann; 2000.
8. Amstutz HE, Horstman LA, Morter RL. Clinical evaluation of the efficacy of *Haemophilus somnus* and *Pasteurella* sp. bacterins. *Bov Pract* 1981;16:106–8.
9. Bateman KG. Efficacy of a *Pasteurella haemolytica* vaccine/bacterial extract in the prevention of bovine respiratory disease in recently shipped feedlot calves. *Can Vet J* 1988;29:838–9.
10. Bechtol DT, Jones GF. Can a *Pasteurella* vaccine prevent respiratory disease in calves in a backgrounding lot? *Vet Med* 1996;91:1042–5.
11. Frank GH, Briggs RE, Duff GC, et al. Effects of vaccination prior to transit and administration of florfenicol at time of arrival in a feedlot on the health of transported calves and detection of *Mannheimia haemolytica* in nasal secretions. *Am J Vet Res* 2002;63:251–6.
12. Ives S, Drouillard J, Anderson D, et al. Comparison of morbidity and performance among stressed feeder calves following vaccination with PYRAMID MLV 4 or PYRAMID 4 + PRESPONSE SQ. Kansas State University, Cattlemen's Day, Report of Progress 1999;831:126–9.
13. Jim K, Guichon T, Shaw G. Protecting feedlot calves from pneumonic pasteurellosis. *Vet Med* 1988;83:1084–7.
14. Loan RW, Tigges MG, Purdy CW. A tissue culture-derived *Pasteurella haemolytica* vaccine. *Bov Pract* 1989;24:22–4.
15. MacGregor S, Smith D, Perino LJ, et al. An evaluation of the effectiveness of a commercial *Mannheimia (Pasteurella) haemolytica* vaccine in a commercial feedlot. *Bov Pract* 2003;37:78–82.
16. Malcolm-Callis KJ, Galyean ML, Duff GC. Effects of dietary supplemental protein source and a *Pasteurella haemolytica* toxoid on performance and health of newly received calves. *Agri Pract* 1994;15:22–8.
17. McLean GS, Smith RA, Gill RA, et al. An evaluation of an inactivated, leukotoxin-rich, cell-free *Pasteurella haemolytica* vaccine for prevention of undifferentiated bovine respiratory disease. *Okla State Univ Anim Sci Res Rep MP-129* 1990:135–40.
18. Purdy CW, Livingston CW, Frank GH, et al. A live *Pasteurella haemolytica* vaccine efficacy trial. *J Am Vet Med Assoc* 1986;188:589–91.
19. Smith RA, Gill DR, Hicks RB. Improving the performance of stocker and feedlot calves with a live *Pasteurella haemolytica* vaccine. *Vet Med* 1986;81:978–81.
20. Thorlakson B, Martin W, Peters D. A field trial to evaluate the efficacy of a commercial *Pasteurella haemolytica* bacterial extract in preventing bovine respiratory disease. *Can Vet J* 1990;31:573–9.
21. Van Donkersgoed J, Schumann FJ, Harland RJ, et al. The effect of route and dosage of immunization on the serological response to a *Pasteurella haemolytica* and *Haemophilus somnus* vaccine in feedlot calves. *Can Vet J* 1993;34:731–5.
22. Morter RL, Amstutz HE. Evaluating the efficacy of a *Haemophilus somnus* bacterin in a controlled field trial. *Bov Pract* 1983;18:82–3.
23. Bechtol DT, Ballinger RT, Sharp AJ. Field trial of a *Pasteurella haemolytica* toxoid administered at spring branding and in the feedlot. *Agri-Pract* 1991;12:6–14.
24. Bennett BW. Efficacy of *Pasteurella* bacterins for yearling feedlot cattle. *Bov Pract* 1982;3:26–30.
25. Martin W, Acres S, Janzen E, et al. A field trial of preshipment vaccination of calves. *Can Vet J* 1984;25:145–7.
26. Ribble CS, Jim GK, Janzen ED. Efficacy of immunization of feedlot calves with a commercial *Haemophilus somnus* bacterin. *Can J Vet Res* 1988;52:191–8.

27. Smith CK, Davidson JN, Henry CW Jr. Evaluating a live vaccine for *Pasteurella haemolytica* in dairy calves. *Vet Med* 1985;80:78–88.
28. Confer AW, Fulton RW. Evaluation of *Pasteurella* and *Haemophilus* vaccines. *Proc 27th Conv AABP*. Pittsburgh (PA):American Association of Bovine Practitioners; 1995. p. 136–41.
29. Confer AW, Ayalew S, Panciera RJ, et al. Immunogenicity of recombinant *Mannheimia haemolytica* serotype 1 outer membrane protein PlpE and augmentation of a commercial vaccine. *Vaccine* 2003;21/22:2821–9.
30. Aubry P, Warnick LD, Guard CL, et al. Health and performance of young dairy calves vaccinated with a modified-live *Mannheimia haemolytica* and *Pasteurella multocida* vaccine. *J Am Vet Med Assoc* 2001;219:1739–42.
31. Van Donkersgoed J, Potter AA, Mollison B, et al. The effect of a combined *Pasteurella haemolytica* and *Haemophilus somnus* vaccine and a modified-live bovine respiratory syncytial virus vaccine against enzootic pneumonia in young beef calves. *Can Vet J* 1994;35:239–41.
32. Blanchard-Channell MT, Ashfaq MK, Kadel WL. Efficacy of streptomycin-dependent, live *Pasteurella haemolytica* vaccine against challenge exposure to *Pasteurella haemolytica* in cattle. *Am J Vet Res* 1987;48:637–42.
33. Cardella MA, Adviento MA, Nervig RM. Vaccination studies against experimental bovine *Pasteurella* pneumonia. *Can J Vet Res* 1987;51:204–11.
34. Catt DM, Chengappa MM, Kadel WL, et al. Preliminary studies with a live streptomycin-dependent *Pasteurella multocida* and *Pasteurella haemolytica* vaccine for the prevention of bovine pneumonic pasteurellosis. *Can J Comp Med* 1985;49:366–71.
35. Chengappa MM, McLaughlin BG, Craft DL. Bovine pneumonic pasteurellosis: efficacy testing a live vaccine. *Vet Med* 1998;83:837–40.
36. Conlon JAR, Gallo GF, Shewen PE, et al. Comparison of protection of experimentally challenged cattle vaccinated once or twice with a *Pasteurella haemolytica* bacterial extract vaccine. *Can J Vet Res* 1995;59:179–82.
37. DeBey BM, Roth JA, Brogden KA, et al. *In vitro* lymphocyte proliferative responses and gamma-interferon production as measures of cell-mediated immunity of cattle exposed to *Pasteurella haemolytica*. *Can J Vet Res* 1996;60:263–70.
38. Shewen PE, Wilkie BN. Vaccination of calves with leukotoxic culture supernatant from *Pasteurella haemolytica*. *Can J Vet Res* 1988;52:30–6.
39. Shewen PE, Lee CW, Perets A, et al. Efficacy of recombinant sialoglycoprotease in protection of cattle against pneumonic challenge with *Mannheimia (Pasteurella) haemolytica* A1. *Vaccine* 2003;21:1901–6.
40. Shewen PE, Sharp A, Wilkie BN. Efficacy testing a *Pasteurella haemolytica* extract vaccine. *Vet Med* 1988;83:1078–83.
41. Srinand S, Mahewsaran SK, Anees TR, et al. Evaluation of efficacy of three commercial vaccines against experimental bovine pneumonic pasteurellosis. *Vet Microsc* 1996;52:81–9.
42. Berghaus LJ, Corbeil LB, Berghaus RD, et al. Effects of dual vaccination for bovine respiratory syncytial virus and *Haemophilus somnus* on immune responses. *Vaccine* 2006;24:6018–27.
43. Cairns R, Chu HJ, Chavez LG, et al. Efficacy of an outer membrane complex *Haemophilus somnus* bacterin in preventing symptoms following *Haemophilus somnus* challenge. *Agri-Pract* 1993;14:35–7.
44. Groom SC, Little PB. Effects of vaccination of calves against induced *Haemophilus somnus* pneumonia. *Am J Vet Res* 1988;49:793–800.
45. Review Manager (RevMan), Version 5.1. Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration, 2011.

Appendix 1 Articles reviewed for evaluation of the effectiveness of commercially available vaccines against <i>M haemolytica</i> , <i>P multocida</i> , and/or <i>H somni</i> in feedlot cattle using natural disease challenge with cumulative morbidity risk and/or cumulative mortality risk reported as an outcome			
Reference	Study description	Vaccine	RR (95% CI)
<i>M haemolytica</i> and <i>M haemolytica</i> + <i>P multocida</i> vaccine studies			
Amstutz et al, Bov Pract, 1981 ⁸	Random allocation of beef heifers vaccinated at feedlot arrival and 21 days later	<i>M haemolytica</i> + <i>P multocida</i> bacterin/ 2 IM doses	BRD morbidity: RR = 1.14 (0.78–1.67) Tx: 38/108 Control: 33/107 BRD mortality: RR = not calculated Tx: 1/108 Control: 2/107
Bateman, Can Vet J, 1988 ⁹	Randomized controlled trial allocation using beef calves (average, 255–268 kg) housed at 2 different locations vaccinated at feedlot arrival	Inactivated <i>M haemolytica</i> , bacteria-free extract with leukotoxin and bacterial surface subunit antigens/1 IM dose	BRD morbidity: RR = 0.89 (0.58–1.30) Tx: 29/90 Control: 32/86 Crude mortality: RR = not calculated Tx: 0/90 Control: 0/86
Bechtol & Jones, Vet Med, 1996 ¹⁰	Randomized block (paired sequentially by processing order) allocation using beef heifers (160–170 kg) vaccinated at spring arrival to 45-day backgrounding lot	Avirulent live culture of <i>M haemolytica</i> and <i>P multocida</i> /1 IM dose	Crude morbidity: RR = 0.81 (0.66–1.00) Tx: 100/295 Control: 124/296 Crude mortality: RR = 1.25 (0.34–4.62) Tx: 5/295 Control: 4/296 (some pens were mass medicated)
Bechtol & Jones, Vet Med, 1996 ¹⁰	Randomized block (paired sequentially by processing order) allocation using beef heifers (160–170 kg) vaccinated at July arrival to 45-day backgrounding lot	Avirulent live culture of <i>M haemolytica</i> and <i>P multocida</i> /1 IM dose	Crude morbidity: RR = 0.85(0.69–1.05) Tx: 78/161 Control: 91/160 Crude mortality: RR = 0.99 (0.14–6.97) Tx: 2/161 Control: 2/160 (some pens were mass medicated)

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Appendix 1 (continued)			
Reference	Study description	Vaccine	RR (95% CI)
Frank et al, AJVR, 2002 ¹¹	Systematic (every-other-steer) allocation using 170–230 kg steers vaccinated at order buyer premises prior to transit to a feedlot	Bacterin-toxoid (chemically inactivated culture of multiple isolate of <i>M haemolytica</i>)/1 IM dose	BRD morbidity: RR = 0.95 (0.72–1.25) Tx: 37/60 Control: 39/60
Ives et al, KSU Cattleman's Day, 1999 ¹²	Systematic (every-other-heifer) allocation using beef heifers (average, 227 kg) vaccinated at feedlot arrival	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/1 IM dose	BRD morbidity: RR = 0.80 (0.62–1.04) Tx: 60/162 Control: 75/162 BRD mortality + chronic animals RR = 0.60 (0.22–1.61) Tx: 6/162 Control: 10/162
Jim et al, Vet Med, 1988 ¹³	Systematic allocation using cattle vaccinated at feedlot arrival and again 1–5 days later	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses	BRD morbidity: RR = 0.95 (0.89–1.03) Tx: 444/781 Control: 766/1291
Loan et al, Bov Pract, 1989 ¹⁴	Unknown allocation of cattle vaccinated at feedlot arrival and 28 days earlier	Bacterin (tissue culture-derived <i>M haemolytica</i> bacterin)/2 IM doses	BRD morbidity: RR = 0.30 (0.09–1.03) Tx: 3/50 Control: 10/50
MacGregor et al, Bov Pract, 2003 ¹⁵	Systematic (every-other-one) allocation using cattle vaccinated at feedlot arrival	Bacterin-toxoid (chemically inactivated culture of multiple isolate of <i>M haemolytica</i>)/1 dose	BRD morbidity: RR = 0.93 (0.83–1.04) Tx: 447/1652 Control: 480/1652 Crude mortality: RR = 0.69 (0.44–1.06) Tx: 33/1652 Control: 48/1652 BRD mortality: RR = 0.71 (0.41–1.22) Tx: 22/1652 Control: 31/1652

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Appendix 1 (continued)	Study description	Vaccine	RR (95% CI)
Malcolm-Callis et al, Agri-Pract, 1986 ¹⁶	Unknown allocation of cattle vaccinated at feedlot arrival and 14 days later	Inactivated <i>M haemolytica</i> , bacteria-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses	BRD morbidity: RR = 0.56 (0.34–0.92) Tx: 16/60 Control: 28/59 Crude mortality: RR = 0.33 (0.04–3.06) Tx: 1/60 Control: 3/59
McLean et al, Oklahoma State Univ Animal Science Research Report, 1990 ¹⁷	Random allocation of cattle vaccinated prior to transit to a feedlot and 7 days later	Inactivated <i>M haemolytica</i> , bacteria-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses	BRD morbidity: RR = 0.84 (0.72–1.00) Tx: 89/147 Control: 109/152 Crude mortality: RR = 0.66 (0.26–1.65) Tx: 7/147 Control: 11/152
McLean et al, Oklahoma State Univ Animal Science Research Report, 1990 ¹⁷	Random allocation of cattle vaccinated at feedlot arrival and 7 days later	Inactivated <i>M haemolytica</i> , bacteria-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses	BRD morbidity: RR = 1.01 (0.88–1.16) Tx: 116/160 Control: 109/152 Crude mortality: RR = 1.21 (0.57–2.58) Tx: 14/160 Control: 11/152
Purdy et al, JAVMA, 1986 ¹⁸	Unknown allocation of calves from a single ranch vaccinated 14 days prior to transit to an order-buyer where they remained for 6 days in contact with other cattle before being transported to a feedlot	Live culture of <i>M haemolytica</i> /1 ID dose	BRD morbidity: RR = 0.94 (0.80–1.11) Tx: 34/41 Control: 52/59 Crude mortality: RR = 0.48 (0.10–2.26) Tx: 2/41 Control: 6/59

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Appendix 1 (continued)			
Reference	Study description	Vaccine	RR (95% CI)
Smith et al, Vet Med, 1986 ¹⁹	Random allocation of steers and bulls vaccinated at feedlot arrival	Live culture of <i>M haemolytica</i> /1 ID dose	Crude morbidity: RR = 0.77 (0.57–1.04) Tx: 55/218 Control: 73/223 Crude mortality: RR = not calculated Tx: 0/218 Control: 1/223
Thorlakson et al, Can Vet J, 1990 ²⁰	Systematic allocation of 6- to 8-month-old cattle vaccinated 21 days prior to transit at ranch of origin (not vaccinated at feedlot)	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/1 IM dose	Crude morbidity: RR = 1.02 (0.84–1.23) Tx: 122/284 Control: 122/289 BRD mortality: RR = not calculated Tx: 5/284 Control: 1/289
Thorlakson et al, Can Vet J, 1990 ²⁰	Systematic allocation of 6- to 8-month-old ranch-fresh cattle vaccinated at feedlot arrival	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/1 IM dose	Crude morbidity: RR = 1.09 (0.91–1.31) Tx: 134/291 Control: 122/289 BRD mortality: RR = not calculated Tx: 0/291 Control: 1/289
Thorlakson et al, Can Vet J, 1990 ²⁰	Systematic allocation of 6- to 8-month-old cattle vaccinated at ranch of origin 21 days prior to transit and at feedlot arrival	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses (arrival and 21 days prior)	Crude morbidity: RR = 0.92 (0.75–1.12) Tx: 107/276 Control: 122/289 BRD mortality: RR = not calculated Tx: 2/276 Control: 1/289

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Appendix 1 (continued)			
Reference	Study description	Vaccine	RR (95% CI)
Thorlakson et al, Can Vet J, 1990 ²⁰	Systematic allocation of auction derived cattle vaccinated at feedlot arrival	Inactivated <i>M haemolytica</i> , bacterial-free extract with leukotoxin and bacterial surface subunit antigens/1 IM dose	Crude morbidity: RR = 0.96 (0.88–1.05) Tx: 259/363 Control: 274/368 BRD mortality: RR = 0.68 (0.28–1.63) Tx: 8/363 Control: 12/368
<i>M. haemolytica</i> + <i>H somni</i> vaccine studies			
Van Donkersgoed et al, Can Vet J, 1993 ²¹	Random allocation of steers (average, 237 kg) vaccinated at feedlot arrival or at arrival and 14 days later	Genetically attenuated leukotoxin of <i>M haemolytica</i> combined with bacterial extracts of <i>M haemolytica</i> + <i>H somni</i> /1 or 2 SQ or IM doses	BRD morbidity: RR = 0.37 (0.24–0.56) Tx: 29/198 Control: 41/103 BRD mortality: RR = NA Tx: 0/198 Control: 2/103
<i>H somni</i> vaccine studies			
Amstutz et al, Bov Pract, 1981 ⁸	Random allocation of beef heifers vaccinated at feedlot arrival and 21 days later	<i>H somni</i> bacterin/2 IM doses	BRD morbidity: RR = 0.64 (0.40–1.03) Tx: 21/106 Control: 33/107 BRD mortality: RR = NA Tx: 3/106 Control: 2/107
Morter & Amstutz, Bov Pract, 1983 ²²	Random allocation of crossbred steers vaccinated at feedlot arrival	<i>H somni</i> bacterin/1 IM dose	BRD morbidity: RR = 1.44 (1.03–2.03) Tx: 49/102 Control: 34/102
Morter & Amstutz, Bov Pract, 1983 ²²	Random allocation of crossbred steers vaccinated at feedlot arrival and 21 days later	<i>H somni</i> bacterin/2 IM doses	BRD morbidity: RR = 0.91 (0.61–1.36) Tx: 31/102 Control: 34/102

Appendix 2

Articles reviewed for evaluation of the effectiveness of commercially available vaccines against *M haemolytica*, *P multocida*, and/or *H somni* in feedlot cattle using an induced disease model with lung lesions or other measures of disease severity reported as an outcome

Reference	Study design	Vaccine	Outcome
	<i>M. haemolytica</i> vaccine studies		
Confer & Fulton, Bovine Proceedings, 1994 ²⁸	Unknown allocation of 136–205 kg beef calves vaccinated twice at 21 day interval prior to transthoracic <i>M haemolytica</i> challenge 14 days after last vaccination	<i>M haemolytica</i> bacterin-solubilized surface antigens	25% of control calves died and surviving cattle developed moderate to severe pneumonia while vaccinated cattle had transient clinical signs of BRD and no deaths. Pulmonary lesions for vaccinates were 64.5%–71.4% less than for controls
Confer & Fulton, Bovine Proceedings, 1994 ²⁸	Unknown allocation of calves vaccinated once prior to transthoracic <i>M haemolytica</i> challenge	<i>M haemolytica</i> bacterin-toxoid (chemically inactivated culture of multiple isolate of <i>M haemolytica</i>)	80% of control cattle and 10% of vaccinated cattle died following challenge. All control calves had severe pneumonia. The surviving vaccinated cattle had transient clinical signs of BRD. Pulmonary lesions for vaccinates were 52.5%–53% less than for controls
Confer et al, Vaccine, 2003 ²⁹	Unknown allocation of weaned beef steers vaccinated once prior to transthoracic challenge with 3.0×10^9 CFU <i>M haemolytica</i> 24 days later	Inactivated <i>M haemolytica</i> , bacteria-free extract with leukotoxin and bacterial surface subunit antigens/2 IM doses	At necropsy, mean lung lesion scores were 7.9 ± 3.6 for nonvaccinated controls and 3.0 ± 1.3 for vaccinates (62.0% reduction in lesion score)
Loan et al, Bov Pract, 1989 ¹⁴	Unknown allocation of 182–227 kg cattle vaccinated twice at 28-day interval and challenged via transthoracic inoculation with <i>M haemolytica</i> 7 days after last vaccination	Bacterin (tissue culture-derived <i>M haemolytica</i> bacterin)/2 IM doses	More vaccinated calves survived 4 days post challenge than controls (14/18 vs 0/18) and at necropsy 4 days post challenge, control calves had lung lesions that averaged 831 cm ³ while vaccinated calf lung lesions averaged 58 cm ³
Loan et al, Bov Pract, 1989 ¹⁴	Unknown allocation of calves vaccinated at 1–4 month of age and returned to their dams, vaccinated again 3 months later and challenged via transthoracic inoculation with <i>M haemolytica</i> 7 days after last vaccination	Bacterin (tissue culture-derived <i>M haemolytica</i> bacterin)/2 IM doses	More vaccinated calves survived 4 days post challenge than controls (8/8 vs 1/8)

Appendix 3 Articles reviewed for the evaluation of effectiveness of commercially available vaccines against <i>M haemolytica</i> , <i>P multocida</i> , and/or <i>H somni</i> in dairy or beef calves using natural disease challenge with cumulative morbidity risk and/or cumulative mortality risk reported as an outcome			
Reference	Study description	Vaccine	RR (95% CI)
<i>M. haemolytica</i> and <i>M haemolytica</i> + <i>P multocida</i> vaccine studies			
Aubry et al, JAVMA, 2001 ³⁰	Paired (sequentially by birth date) allocation of Holstein heifer calves first vaccinated at 14 and 20 days of age and again 14 days later	Avirulent live culture of <i>M haemolytica</i> and <i>P multocida</i> /2 IM doses	BRD morbidity: RR = 1.00 (0.64–1.56) Tx: 32/179 Control: 32/179 Crude mortality: RR = 0.86 (0.29–2.50) Tx: 6/179 Control: 7/179
Smith et al, Vet Med, 1985 ²⁷	Unknown allocation of dairy bull calves vaccinated once at 2 weeks of age (1984 vaccinates)	Live culture of <i>M haemolytica</i> /1 ID dose	BRD morbidity: RR = 0.55 (0.41–0.74) Tx: 31/73 Control: 59/77
Smith et al, Vet Med, 1985 ²⁷	Unknown allocation of dairy bull calves vaccinated once at 2 weeks of age (1985 vaccinates)	Live culture of <i>M haemolytica</i> /1 ID dose	BRD morbidity: RR = 0.62 (0.47–0.82) Tx: 34/74 Control: 58/78 Crude mortality: RR = 0.21 (0.05–0.93) Tx: 2/74 Control: 10/78
<i>M. haemolytica</i> + <i>H somni</i> vaccine studies			
Van Donkersgoed et al, Can Vet J, 1994 ³¹	Systematic allocation of beef calves vaccinated at 3 weeks and again at 5 weeks of age	Genetically attenuated leukotoxin of <i>M haemolytica</i> combined with bacterial extracts of <i>M haemolytica</i> + <i>H somni</i> /1 or 2 SQ or IM doses	BRD morbidity: RR = 1.04 (0.51–2.10) Tx: 10/28 Control: 10/29
Van Donkersgoed et al, Can Vet J, 1994 ³¹	Systematic allocation of beef calves vaccinated at 3 weeks and again at 5 weeks of age	Genetically attenuated leukotoxin of <i>M haemolytica</i> combined with bacterial extracts of <i>M haemolytica</i> + <i>H somni</i> /1 or 2 SQ or IM doses	BRD morbidity: RR = 0.44 (0.16–1.25) Tx: 4/26 Control: 10/29